MiR-224 ameliorates inflammation and symptoms in mouse model of allergic rhinitis by targeting CDK9

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Abstract

Objectives: To explore the regulatory effects of microRNA (miRNA)-224 and its potential target gene, cyclin dependent kinase 9 (CDK9), in the pathological process of allergic rhinitis (AR).

Methods: To investigate the role of miR-224 and CDK9, it was screened by bioinformatics prediction software and verified by dual-luciferase reporter assay. The mouse model of AR was established by ovalbumin (OVA). The animal models were intervened with miR-224 agomir, negative control agomir, and saline respectively. The symptoms of sneezing and nasal rubbing were recorded. The expressions of miR224, CDK9, and cytokines in the nasal mucosa of different groups were analyzed by rt-PCR or western blotting. Enzyme-linked immunoassay (ELISA) was used to evaluate the levels of IgE and Histamine (HA) in the serum. The infiltration of inflammatory cells in the nasal mucosa was studied by immunohistochemistry. The expression and distribution of CDK9 in the nasal mucosa of mice were revealed by immunofluorescence.

Results: In the nasal mucosa of the animal models, the level of miR-224 was downregulated, while that of CDK9 was upregulated. The upregulation of miR-224 by miR-224 agomir reduced the frequencies of nasal rubbing and sneezing, the expression of CDK9, the levels of cytokines, and the concentrations of IgE and HA. Moreover, miR-224 appeared to attenuate the infiltration of inflammatory cells and hypersecretion of glands in the nasal mucosa. The expression of CDK9, which was distributed under the mucosa, especially in the submucosa interstitial tissue, was significantly reduced.

Conclusion: MiR-224 affected the pathogenesis of AR by targeting CDK9. It proves that miR-224 could be a novel potential therapeutic target for AR.

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KEYWORDS
rhinitis; allergic; microRNAs; cyclin-dependent kinase 9; genetic therapy

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Introduction

Allergic rhinitis (AR) is a complex allergic disease of rhinology with multigene involvement. One of the most common allergic diseases in the world, the prevalence of AR worldwide is 10% to 40%, and in China it has exceeded 17% with a significant upward trend in recent years. AR seriously affects patients’ quality of life, and at the same time adds a huge medical and socio-economic burden on them. The pathogenesis of AR is complex. A variety of immunoreactive and pro-inflammatory cells and cytokines are thought to be involved in it at present, but no satisfactory treatment for AR is available. The developments in the molecular pathological mechanisms reveal that there are differential expressions of multiple gene sequences in the pathogenesis of AR, involving multiple ribonucleic acids (RNA) such as long-stranded non-coding RNA (lncRNA) and microRNA (miRNA). MiRNAs are a class of small molecule RNAs which was discovered in recent years that affect and regulate cell growth, differentiation, and function. The MiRNAs play an important role in the post-transcriptional regulation of protein-coding gene expression, thereby affecting a range of physiological and pathological processes in the organism. AR is a disease associated with multiple genes, and thereby miRNAs may participate in its development. A previous study showed that compared to the control group, 421 miRNAs were differentially expressed in the nasal mucosa of AR patients, and nine miRNAs in AR group had significant differences with twofold change. A previous study showed that compared to the control group, 421 miRNAs were differentially expressed in the nasal mucosa of AR patients, and nine miRNAs in AR group had significant differences with twofold change. Of these 9 miRNAs, 7 (miR-498, miR187, miR-143, miR-886-3p, miR-224, and miR-767-5p) decreased in expression and 2 (miR-7 and miR-224) increased. MiR-224 is significantly downexpressed in the nasal mucosa of AR patients, suggesting that it may be involved in the regulatory network of genes associated with allergic inflammation, but its mechanism requires further study. In the present study, the possible target genes of miR-224 were predicted by miRNA target gene prediction software and database respectively. The intersection of at least two target gene prediction software was taken as a result, and cell cycle protein-dependent kinase 9 (CDK9) was finally screened as one of the potential target genes of miR-224, and its role in AR was explored.

CDK9 is a member of the serine/threonine kinase family, which is essential in the whole cell cycle and apoptosis. CDK9 is involved in inflammatory response, viral infection, and tumor cell regulation. Th2 lymphocyte cytokines, mast cell, and endothelial cell related cytokines, including IL-4, IL-5, IL-6, IL-13, etc. modulate the immune reaction in AR. CDK9 interacts with the receptor of IL-6 family of cytokines, gp130, and affects the inflammatory reaction. Additionally, the differentiation and apoptosis of Treg cells are both associated with the pathologic process of AR. Study has shown that CDK9 inhibition increases the regulatory T cell population, induces apoptosis in leukocyte subsets PBMCs, and modulates the immune response. Therefore, a change the expression of CDK9 will likely benefit the treatment of human diseases, including allergic diseases. CDK9 has been extensively studied in the field of tumor therapy, and its regulatory mechanisms are still being explored. CDK9 is closely related to the release of various cytokines and the regulation of RNA. Based on the fact that AR is a multi-gene disease with complex pathogenesis, the mechanism of CDK9’s role in it needs to be further explored.

In this experiment, the potential target gene CDK9 of miR-224 was predicted, screened, and validated. Furthermore, we used miR-224 agomir to intervene the AR mouse model to examine the regulatory function of miR-224 on the target gene, and to explore the role of miR-224 and CDK9 in the pathogenesis of AR, which will provide a theoretical basis for future viable gene therapy for AR.

Materials and methods

AR animal model

All animal experiment procedures were carried out according to the Animal Testing Guideline of the Animal Laboratory Centre of our affiliation. C57 mice (male, 8-weeks old) were provided by the Animal Laboratory Centre of our affiliation. The mice were randomly divided into four groups of eight mice: (1) AR group; (2) AR + agomiR224 (miR224 agomir) intervention group; (3) AR + NC-miRNA (negative control miRNA agomir) control group; (4) saline control group. AgomiR224 and NC-miRNA were synthesized by Shanghai Jima Bio. The mice of the first three groups were injected intraperitoneally with OVA (40 μg/kg, Solarbio, Beijing, China) and aluminum hydroxide gel (40 mg/kg, Sigma Aldrich, USA) on day 1, day 7, day 14, and day 21. The saline control group was injected intraperitoneally with saline at the same time. After the last abdominal sensitization, nasal drop (morning) was administered in the morning to the latter three groups of mice as the process of intervention using agomiR224 (20 μL, 1 μg/μL), NC-miRNA (20 μL, 1 μg/μL), and saline (20 μL), respectively. Subsequently, the AR groups of mice were challenged to sensitize by the continuous intranasal drop of OVA (20 μL, 25 mg/mL) for three days (afternoon). The intervention and OVA challenged processes were then repeated thrice more (4 times in total). There was a six hour interval between intervention and challenge (Figure 1). After the last nasal challenge, the numbers of sneezing and nasal rubbing of the mice were recorded during a 10 minute interval period. The mice were anesthetized by intraperitoneal injection of sodium pentobarbital (40mg/kg). Fresh blood from the tail of each mouse was divided into two tubes, and the blood samples were centrifuged and stored at −20°C. After execution of the mice, the nasal cavity of the mice was quickly dissected to obtain the nasal mucosal tissue, which was immediately transferred to −70°C liquid nitrogen for storage. In addition, nasal tissue was fixed in 10% formaldehyde for histopathological testing.

Bioinformatics analysis

Several miRNA target gene prediction software and databases were used, including TargetScan, miRanda, PITA, picTar, miRwalk, miRTarBase, etc. The predicted target
genes of miR-224 were predicted and exported separately through online service sites. After the localization run, the intersections were taken, and the intersections of at least two target gene prediction software were taken as the results.

**Cell culture and miRNA transfection**

Human nasal mucosal epithelial cells (HNECs) were collected from non-chronic nasal-sinusitis patients undergoing nasal endoscopic surgery by gentle brushing in, a method described by Hu et al.\(^\text{13}\) (approved by the ethics committee of our affiliation). HNECs were maintained with Pneumacult\textsuperscript{TM}-Ex Plus medium (Stem Cell Technologies, Vancouver, Canada) in a cell incubator at 37°C, with 5% CO\(_2\). The transfection of each group was performed according to the instructions of Lipofectamine 2000 (Invitrogen Co., USA).

**Dual-luciferase reporter assay**

Dual-luciferase reporter assay was performed to verify the relationship between miR-224 and the target gene CDK9 based on the results of bioinformatics analysis. The wild type CDK9 3'UTR segment (Genbank accession No. NM_001261) was amplified and cloned into the pGL3-Promoter vector (Promega Co., USA). The mutant CDK9 3'-UTR binding site sequence was determined based on hsa-miR-224-5p and CDK9 3'-UTR binding site, which was inserted into the vector pGL3-Promoter to construct pGL3-Promoter-mutCDK9. HNECs were co-transfected with agomiR224 or NC-miRNA and 3'UTR CDK9 (with wild type or mutant miR-224 binding sites) for 48 hours by Lipofectamine 2000, and then assayed with the dual luciferase reporter assay system (Promega, Madison, WI, USA).

**Real-time PCR**

Total RNA was extracted from mouse nasal mucosal tissue according to the Trizol reagent instructions (ABI-7300, Thermo Fisher, USA). After eliminating the DNA from the total RNA, the first strand of cDNA was synthesized using a reverse transcription kit (Fermentas, Canada). The SYBR Green PCR kit (Thermo Fisher, USA) and the ABI-7300 real-time detector were then used for real-time quantitative PCR detection. The sequences of primers specific to the genes tested are listed in Table 1. The reaction procedure was as follows: PCR amplification: 95°C, 10 min; (95°C, 15 Sec; 60°C, 45 Sec) x 40 cycles; melting curve: 95°C, 15 Sec; 60°C, 1 min; 95°C, 15 Sec; 60°C, 15 Sec. Data analysis was performed by ABI Prism 7300 SDS software.

**Western blotting**

Total tissue protein was extracted by RIPA tissue cell rapid lysis fluid (JRDUN Biotechnology, Shanghai). Protein concentration was quantified by BCA protein quantification kit (Thermo Fisher, USA). An aliquot of 50μg protein was added to a 10% polyacrylamide gel. After electrophoresis, the sample was transferred to a polyvinylidene fluoride (PVDF) membrane, and closed with 5% skimmed milk powder at 4°C overnight. The membrane was then left at room temperature for 30 min, followed by incubation with the respective primary antibody at 4°C overnight. After washing, the membrane was incubated with secondary antibody at 37°C for 30 min. The signal was visualized using chemiluminescence.

**Table 1** The primer sequences used in RT-PCR.

<table>
<thead>
<tr>
<th>Genes</th>
<th>Primers sequences (5' to 3')</th>
</tr>
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<tbody>
<tr>
<td>miR-224</td>
<td>Forward CGGCCGTAAAGTCATAGTGTT</td>
</tr>
<tr>
<td></td>
<td>Reverse AGTGCGAGGCAGCCGATT</td>
</tr>
<tr>
<td>CDK9</td>
<td>Forward AGGTGAGCTTGAAAGACCTG</td>
</tr>
<tr>
<td></td>
<td>Reverse AGCGGTTATACGTTGAGG</td>
</tr>
<tr>
<td>IL-4</td>
<td>Forward GTTGTGATCGTCGTCCCTTC</td>
</tr>
<tr>
<td></td>
<td>Reverse GTTTGGCACATCATCTTC</td>
</tr>
<tr>
<td>IL-6</td>
<td>Forward GAGATACACCATCACCACGAC</td>
</tr>
<tr>
<td></td>
<td>Reverse AAAGTGCATACGTGTTGATCATA</td>
</tr>
<tr>
<td>IL-18</td>
<td>Forward CGTTGAGCAGCAACAGACGAC</td>
</tr>
<tr>
<td></td>
<td>Reverse TACAGGGCAGCTACGACC ENG</td>
</tr>
<tr>
<td>GAPDH</td>
<td>Forward CTGCCAGAAACATCATCTCC</td>
</tr>
<tr>
<td></td>
<td>Reverse CTCAAGATGCGCTTGTTCAC</td>
</tr>
<tr>
<td>U6</td>
<td>Forward GCCCTCAGCAGCACCCAGC</td>
</tr>
<tr>
<td></td>
<td>Reverse GGAACGCTTTGACG</td>
</tr>
</tbody>
</table>
temperature for two hours with diluted antibodies against CDK9 (1:5000, Abcam, US) and glyceraldehyde-3-phosphate dehydrogenase (GAPDH) (1:2000, CST, USA). The membrane was then washed and incubated with diluted horseradish peroxidase (HRP)-labeled sheep anti-rabbit antibodies (1:1000, Beyotime, China) at 37°C for one hour. ECL luminescence solution was formulated and added to the front of the film, followed by five minutes of light avoidance in a dark room. The film was then placed into the imaging system for scanning.

ELISA

The levels of HA and IgE in the serum were tested by mouse HA and IgE ELISA kit (Abcam, US). The concentrations of HA and IgE in the measured serum samples were calculated from the OD values of the samples through standard curves.

Histological staining

After the tissue specimens were fixed, they were encased in paraffin and cut into 5-μm-thick sections, and stained with hematoxylin and eosin. The pictures of the slices were taken using a Lycra microscope (magnification 200).

Immunofluorescence staining

After fixation, embedding, and sectioning of the tissue specimens, antigen repair (hyperbaric repair) was performed. The diluted antibody against CDK9 (1:100) was then added, followed by the diluted fluorescent secondary antibody (1:200).

Statistical analysis

All statistical analyses were performed by SPSS version 20 (IBM Corp, Armonk, NY). The data are presented as mean ± standard error. Statistical significance was determined by one-way ANOVA. P < 0.05 was considered statistically significant.

Results

Effect of agomiR-224 on the symptoms of AR

After the last nasal challenge by OVA, the following nasal symptoms were observed in the mice (Figure 2):

The mice in the AR group and AR + NC-miRNA control group showed obvious allergic symptoms such as sneezing and nasal rubbing (p < 0.001). After OVA challenge, compared to the AR group, mice in AR + agomiR224 intervention group showed a significant reduction in the numbers of nasal rubbing (p < 0.05) and sneezing (p < 0.05); There was no significant difference between AR + NC-miRNA control group and AR group, whereas the saline control group showed almost no symptoms of sneezing and nasal rubbing.

Expression of miR-224 in nasal mucosa

Similar to previous studies, the expression of miR-224 is significantly decreased in the nasal mucosa of AR mice. To further reveal the role of miR-224 in AR, we spotted agomiR224 into the nasal cavity of AR mice. The MiR-224 expression was significantly restored in the nasal mucosa of agomiR224-interfered AR mice, indicating successful gene transduction in these mice (Figure 3).

Expressions of cytokines in the nasal mucosa and levels of IgE, HA in the serum of mice

The involvement of multiple immunoreactive cells and cytokines in the pathogenesis of AR led us to investigate whether the upregulation of miR-224 reduces the level of IL-4 (reflecting Th2 cell levels), IL-6 (reflecting the acute phase of the disease), and IL-18 (reflecting the AR sustained state). The results showed that the levels of these cytokines are significantly reduced in the nasal mucosa of

![Figure 2](image.png) Nasal symptoms of mice within 10 minutes after nasal OVA challenge. (A) Count of nasal rubbing. (B) Count of sneezing. NS: saline control group, AR: AR group, AR-miR224: AR-agomiR224 intervention group, AR-NC-miRNA: AR-NC-miRNA (negative control miRNA agomir) control group. *p < 0.05; ***p < 0.001.
reduced the luciferase activity of the 3′-UTR of the CDK9 gene, significantly attenuated the increase of expression in AR mice with agomiR224 intervention, compared to the AR group (Figure 4A-C). The serum levels of IgE and HA were significantly up-regulated in the AR group of mice compared to the saline control group, while the levels of both were significantly reduced in the agomiR224 intervention group (Figure 4D-E).

CDK9 was the target gene of miR-224

CDK9 was selected as one of the miR-224 target genes based on miRNA target gene prediction software and databases (the mating sites are shown in Figure 5A). The 7-bp fragment of the 3′-UTR of the CDK9 gene is complementary to the miR-224 seed sequence (Figure 5A). As shown in Figure 5C, treatment with agomiR224 significantly reduced the luciferase activity of the 3′-UTR of the wild-type pGL3-Promoter-wtCDK9 vector (p < 0.01), while there was no significant difference in the luciferase activity of the mutant. It was verified that CDK9 is a miR-224 target gene, and is down-regulated by miR-224.

Expression and distribution of CDK9 in the nasal mucosa of mice

We further investigated whether miR-224 affected CDK9 expression in AR. The results showed that CDK9 expression in the nasal mucosa of AR mice was significantly higher than that of normal control mice, while intra-nasal dosing of agomiR224 significantly attenuated the increase of expression of CDK9 induced by OVA sensitization (Figure 6A-B). We also investigated the expression of CDK9 in the nasal mucosa of mice of different groups by immunofluorescence. The results showed that in the AR group and the AR-NC-miRNA group, CDK9 was mainly distributed on the mucosal surface, mucosal epithelium, and glandular cells. After the agomiR224 intervention, the expression of CDK9 showed a decrease in all parts of the nasal mucosa of mice, especially in the mucosal epithelial mesenchyme. In saline normal controls, CDK9 expression was relatively low at all mucosal sites (Figure 6C).

Inflammatory infiltration of the nasal mucosa

Mouse nasal mucosal tissue was sectioned and HE stained. On further microscopic examination, a large number of inflammatory cells, including mast cells, macrophages, and eosinophils, were observed under the nasal mucosa in the AR group and the AR-NC-miRNA group. Abundant vacuolated vesicles between columnar epithelial cells of the nasal mucosa were also observed, suggesting a hyperproduction of glands. In the AR-miR224 group and the NS group, the number of inflammatory cells was significantly lower and the columnar epithelial cells were more closely arranged (Figure 7).

Discussion

As a kind of refractory and recurrent disease, AR seriously affects the physical and mental health as well as the quality of life of patients.15,16 MiRNAs have immunomodulatory effects and participate in the differentiation and maturation of T and B lymphocytes, the regulation of immune response, the secretion of inflammatory cytokines, and other immune processes.17 Over the past two decades, miRNAs have emerged as key regulators that may be involved in the pathogenesis of various diseases, including in respiratory allergic diseases such as allergic rhinitis, allergic asthma,18,19 etc. It has been shown that miRNAs can down-regulate the expression of homologous target genes by cleavage of the target gene transcriptome or repression of its translation.20 Since miRNAs are more stable than mRNAs and even some proteins, researchers believe that this stability is more conducive to the regulation of gene expression in clinical diseases, including AR.19,21 For example, miR-199-3p targets Dnmt3a, resulting in hypermethylation of promoter as well as overexpression of signal transducer and activator of transcription 3 (STAT3), ultimately leading to AR symptoms.22 The anti-inflammatory effect of miR-146a on AR may act by inhibiting the toll-like receptor 4 (TLR4)/TNF receptor-associated factor 6 (TRAF6)/nuclear factor kappa-B (NF-κB) signaling pathway.23 Matrilin 2 (MATN2) can be directly targeted by miR-202-5p, and can reverse miR-202-5p-mediated M2 phenotypic polarization in AR.24 Intranasal administration of lentiviral miR-135a in mice modulates mast cell and allergen-induced inflammation by targeting GATA-binding protein 3 (GATA-3).25 The CDK9-related pathway is a key factor in the regulation of mammalian gene expression, which controls several physiological processes such as cell growth, proliferation, survival, and differentiation.26 Researchers found that the inhibition of CDK9 helped to reduce cytokine production against bacterial infections, such as reducing the expression of both IL-6 and Tumor Necrosis Factor α (TNF-α). Their

Figure 3 MiR-224 expression in nasal mucosa of mice by rt-PCR. NS: saline control group, AR: AR group, AR-miR224: AR-agomiR224 intervention group, AR-NC-miRNA: AR+NC-miRNA (negative control miRNA agomir) control group. *p < 0.05; **p < 0.01.
MiR-224 alleviates AR response by targeting CDK9

In the present study, we explored for the first time, the effect of miR-224 on AR-related allergic reactions induced by OVA in a mouse model. We found that miR-224 ameliorated allergic inflammation in AR mice by targeting CDK9 and inhibiting its expression. These results provide a link between miR-224 and CDK9 in the pathogenesis of AR. Consistent with previous studies, we found reduced expression of miR-224 in the nasal mucosa of AR model mice, further suggesting that miR-224 may negatively regulate the allergic response of AR. To avoid complications that may be caused by intravenous infusion on mice, we took the nasal drop of agomiR224 and studied the direct effect of miR-224 on nasal mucosa. As expected, miR-224 expression was significantly restored after the nasal drip with agomiR224; moreover, nasal rubbing and sneezing symptoms, concentrations of IgE and HA in serum were significantly reduced in mice in the agomiR224 intervention group compared to the AR group. At the same time, the histological analysis also revealed that columnar epithelial cells in the nasal mucosa of AR mice intervened with agomiR224 were closely aligned, and the number of inflammatory cells was significantly reduced. This suggested that agomiR224 actually relieved allergic symptoms in AR mice.

Study also confirmed that CDK9 is involved in the innate immune response. In the present study, we explored for the first time, the effect of miR-224 on AR-related allergic reactions induced by OVA in a mouse model. We found that miR-224 ameliorated allergic inflammation in AR mice by targeting CDK9 and inhibiting its expression. These results provide a link between miR-224 and CDK9 in the pathogenesis of AR. Consistent with previous studies, we found reduced expression of miR-224 in the nasal mucosa of AR model mice, further suggesting that miR-224 may negatively regulate the allergic response of AR. To avoid complications that may be caused by intravenous infusion on mice, we took the nasal drop of agomiR224 and studied the direct effect of miR-224 on nasal mucosa. As expected, miR-224 expression was significantly restored after the nasal drip with agomiR224; moreover, nasal rubbing and sneezing symptoms, concentrations of IgE and HA in serum were significantly reduced in mice in the agomiR224 intervention group compared to the AR group. At the same time, the histological analysis also revealed that columnar epithelial cells in the nasal mucosa of AR mice intervened with agomiR224 were closely aligned, and the number of inflammatory cells was significantly reduced. This suggested that agomiR224 actually relieved allergic symptoms in AR mice.

Figure 4  Expression of cytokines in the nasal mucosa of mice and levels of IgE, HA in the serum of mice. (A)–(C): Rt-PCR was used to analyze the expressions of IL-4, IL-6 and IL-18. (D)–(E): The concentrations of IgE and HA were measured by ELISA. NS: saline control group, AR: AR group, AR-miR224: AR+ agomiR224 intervention group, AR-NC-miRNA: AR+NC-miRNA (negative control miRNA agomir) control group. **p < 0.01; ***p < 0.001.
In conclusion, our present study demonstrated that upregulation of miR-224 effectively ameliorated pathological changes in AR by inhibiting the inflammatory response, including reducing IgE and histamine concentrations, inflammatory cell infiltration, and cytokine levels. Notably, we demonstrated that miR-224 directly targeted CDK9 and inhibited its expression. The underlying mechanisms of its specific signaling pathway need to be studied in depth. Moreover, we reported a novel approach to treat allergic rhinitis by agomiRNA interventions, in which synthetic miR-224 agomir nasal drops can reduce the symptoms of AR by reducing the local allergic inflammatory response in the nasal mucosa through inhibition of the target gene CDK9. This approach might become a potential gene therapy for nasal diseases in the future.

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Potential conflict of interest

None disclosed.

References


4. Zhang J, Li G, Ye X. Cyclin T1/CDK9 interacts with influenza A virus polymerase and facilitates its association with cellular


